





Breeding for perfection

Journey towards understanding the genetic basis of the perfect pearl

Advanced animal breeding in Aquaculture: Using genome-wide molecular breeding values for rapid animal improvement in the silver-lipped pearl oyster



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- *Pinctada maxima* is the primary species of Australia and Indonesia's pearling industry
 - \circ ~ 40% global pearl production value
- Only oyster to produce highly-sought large silver or gold pearls (>15mm)
- Large variation in pearl quality
 - 5% of pearl > 80-90% revenue
- Advancements in hatchery techniques
 - production of hatchery seed stock
 - \circ $\,$ initiation of selection breeding programs $\,$





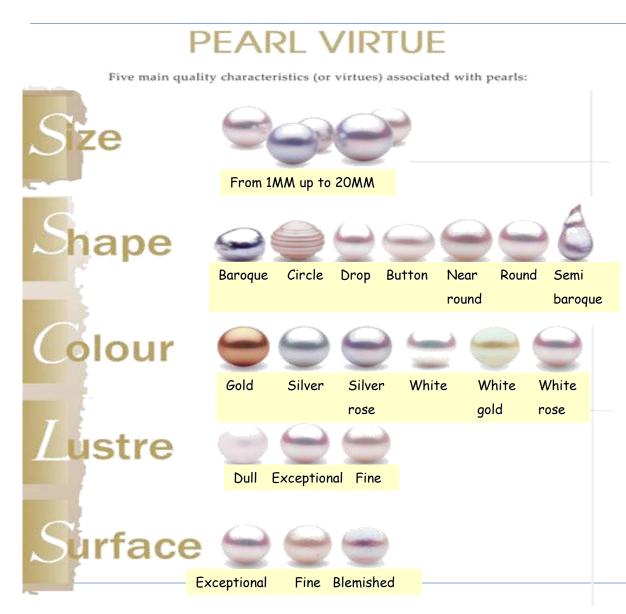


Cultured pearl production complex

<u>Saibo</u> Donor oyster Host oyster Single pearl = 2 animals/genomes 2 years host + 2 years pearl formation **Nucleus**

Pearl trait variability





Gold colour quality



- •12mm Yellow round pearl "A" – US \$30
- •12mm Gold round pearl "AAAA" - US \$2080

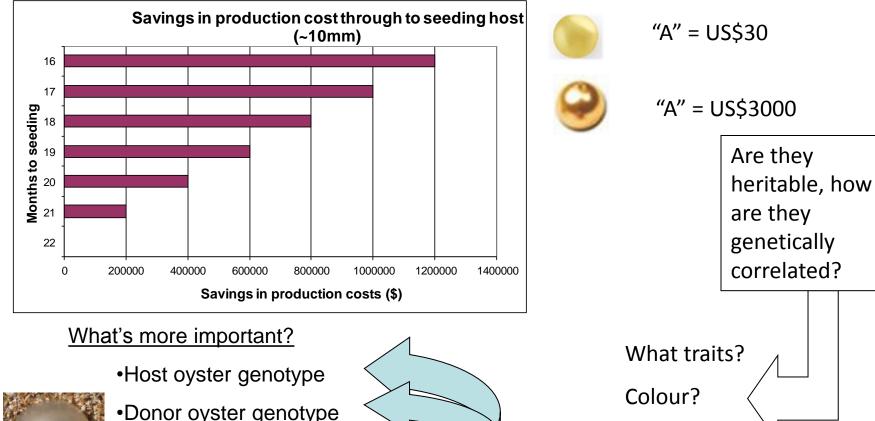
Productivity and Commercial value SIAMES COOK

- Productivity/profitability in industry linked to
 - Product quality
 - Product uniformity
 - Farm cost reduction
- Achieved through
 - Husbandry/Breeding
 - Nutrition
 - Environmental manipulation
 - o Genetic improvement





Productivity and Commercial value JAMES COOK USTRALIA

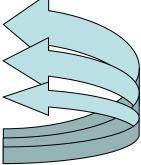




Donor oyster genotype

•Host/Donor genotype interaction

•Environment....

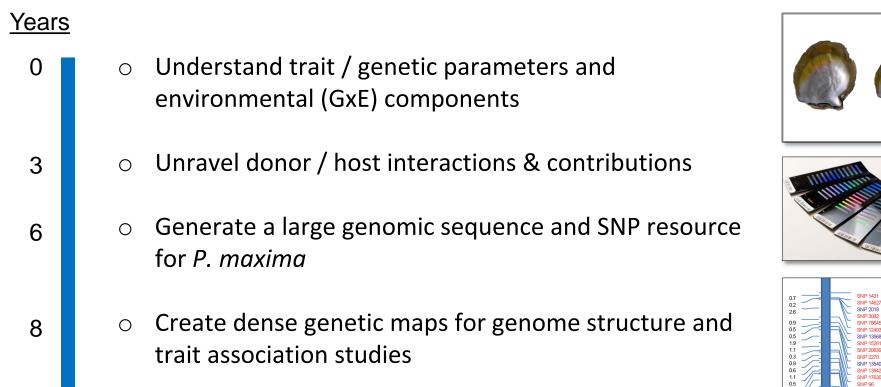


Lustre?

Nuclei Retention?

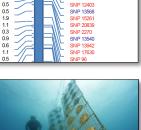
P. maxima Program Objectives





 Integrate genomic selection breeding program based on commercial farm datasets and resources

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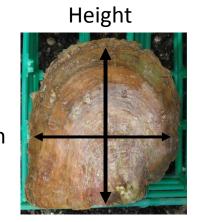
P. maxima genetic parameters



<u>Genetic Parameters – Oyster shell growth traits</u>

Table 1 Heritability ± standard deviation (bold diagonal), genetic (upper diagonal) for oyster shell growth and pearl quality traits. ** indicates the correlation is significant at the 0.01 level (2-tailed), and * indicates the correlation is significant at the 0.05 level (2-tailed).

	Oyster Shell Growth						
Heritabilities ±SD	Length (APM)	Height (DVM)	Width (SW)	Wet weight (WW)			
Length (APM)	0.328 ± (0.128)	0.904**	0.237**	0.852**			
Height (DVM)	-	0.286 ± (0.126)	0.258**	0.882**			
Width (SW)	-	-	0.128 ± (0.081)	0.249**			
Wet weight (WW)	-	-	-	0.183 ± (0.103)			



Width



Length

P. maxima genetic parameters



<u>Genetic Parameters – Pearl quality traits</u>

Table 2. Heritability ± standard deviation (bold diagonal), genetic (upper diagonal) and phenotypic correlations (below diagonal) for pearl quality traits. ** indicates the correlation is significant at the 0.01 level (2-tailed), and * indicates the correlation is significant at the 0.05 level (2-tailed).

	Pearl Quality							
Heritabilities ±SD	Size	Weight	Surface	Colour (SW.O.G)	Colour (G.O)	Colour (S.O)	Colour (W.O)	Colour (SW.O)
Size	0.144 ± (0.026)	0.957**	-0.110*	0.044	-0.031	0.014	0.049	0.049
Weight	0.967**	0.161 ± (0.026)	-0.114*	0.017	-0.001	0.002	0.032	0.029
Surface	-0.194**	-0.195**	0.265 ± (0.031)	0.087	-0.055	0.065	0.083	0.104*
Colour (SW.O.G)	-0.010	-0.027	0.117*	0.363 ± (0.031)	-0.911**	0.457**	0.789**	0.910**
Colour (G.O)	0.012	0.022	-0.090	-0.920**	0.338 ± (0.030)	-0.347**	-0.562**	-0.658**
Colour (S.O)	0.045	0.017	0.122*	0.454**	-0.366**	0.138 ± (0.026)	0.005	0.486**
Colour (W.O)	-0.014	-0.031	0.091	0.799**	-0.593**	0.025	0.235 ± (0.030)	0.876**
Colour (SW.O)	-0.005	-0.028	0.131*	0.907**	-0.687**	0.480**	0.879**	0.270 ± (0.029)



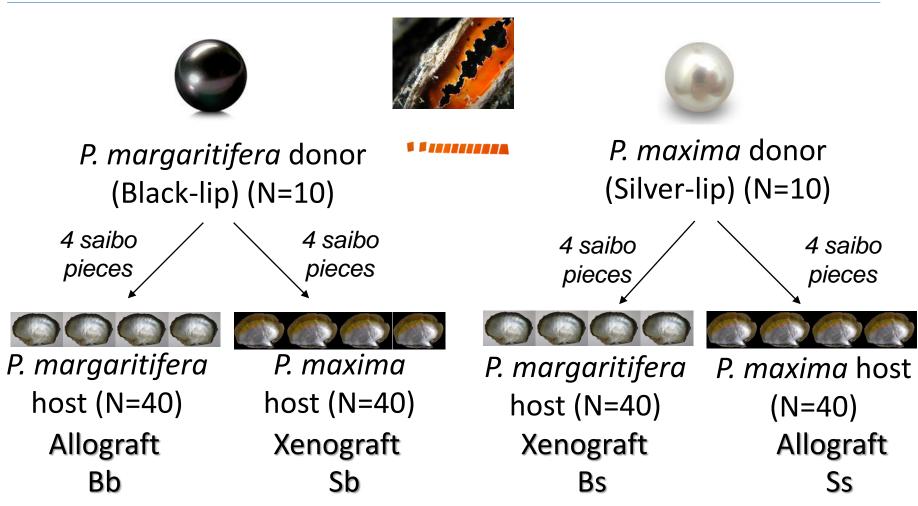




Circle Semi-Baroqu

So what is more important host or donor?



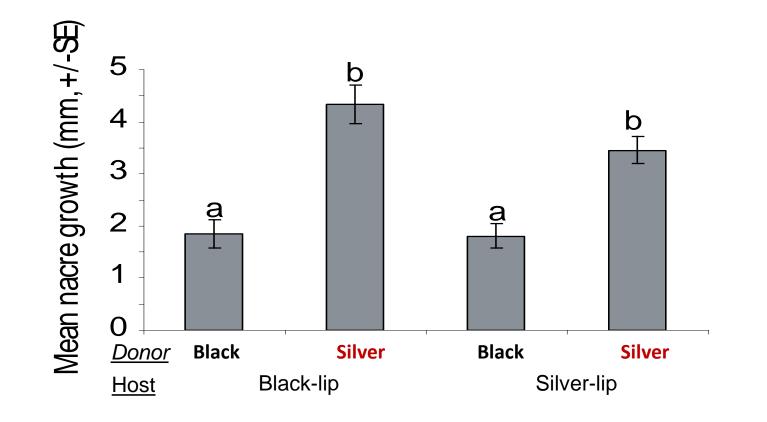


• Nuclei retention and pearl quality recorded 14 months post operation.

Pearl nacre growth



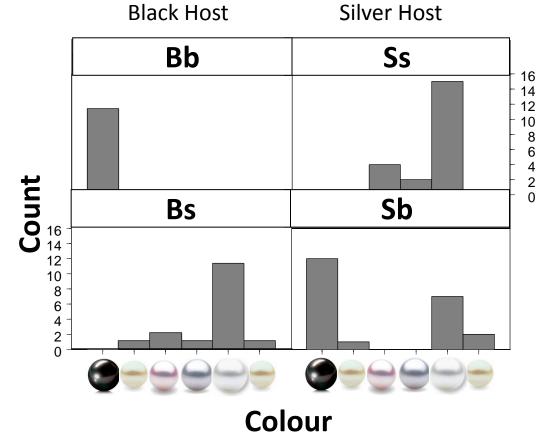
 \circ Pearl growth strongly influenced by donor oyster (P < 0.001)







 Pearl colour was strongly influenced by the donor oyster species (P<0.001).

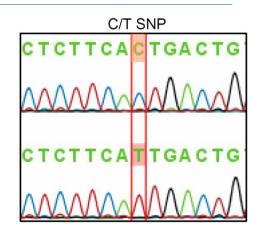


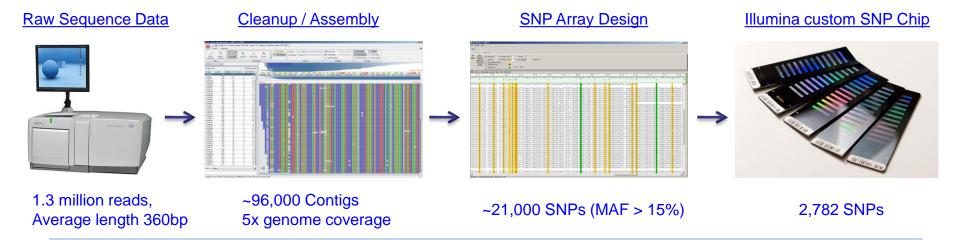
P. maxima genetic resources

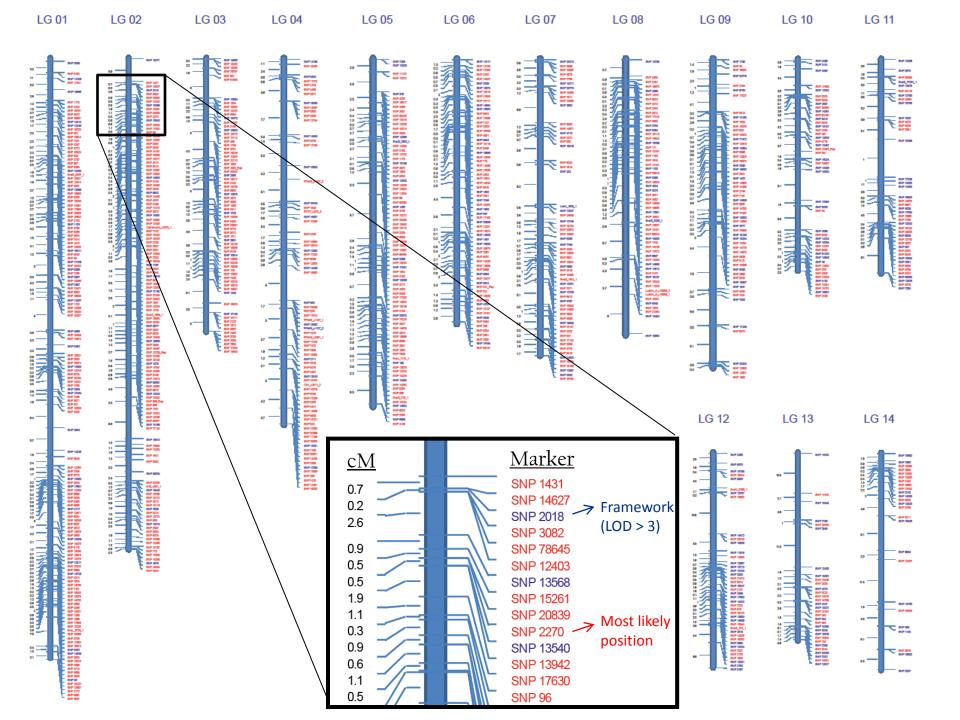


<u>Genome-wide sequencing / SNP chip development</u>

- Mantle / pearl sac tissue RNA to cDNA
- Next-generation sequencing (Roche 454, HiSeq)
- Sequence cleanup & de-novo EST assembly
- Random SNP discovery (MAF & No. reads > 10)
- Custom SNP array designed (many Type I markers)

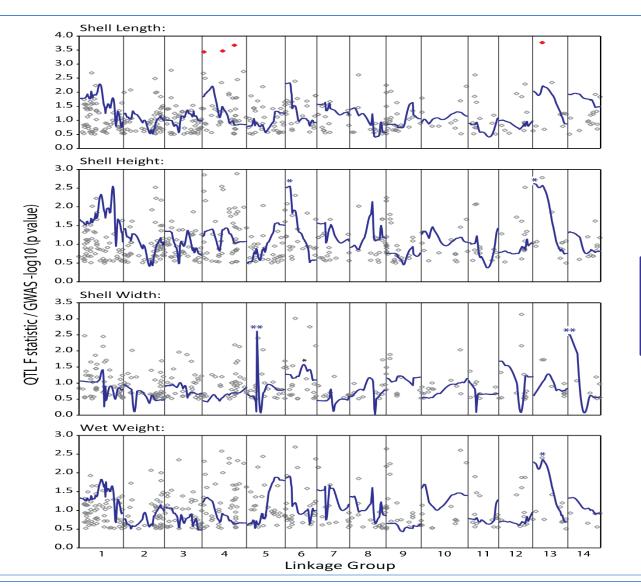






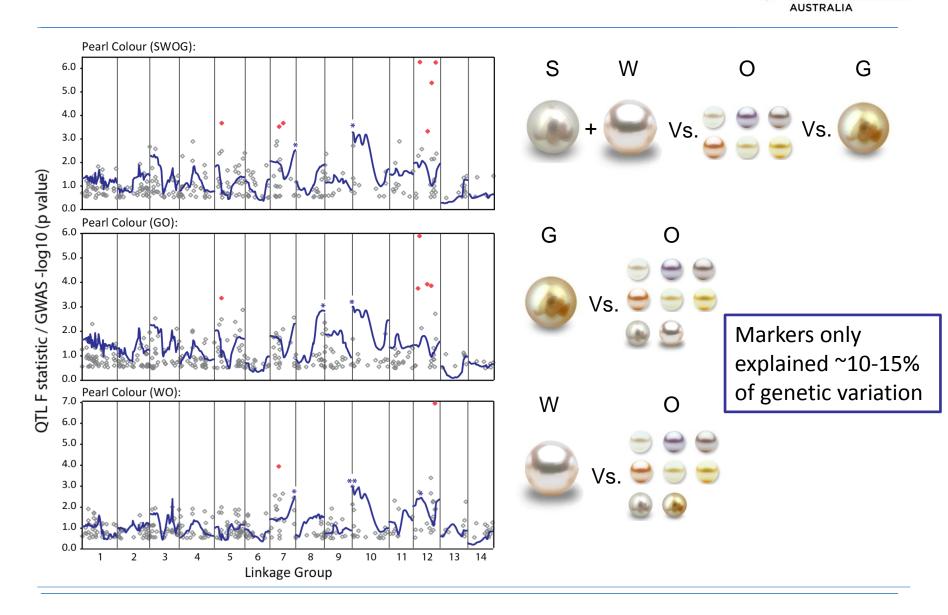
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				• Avera	age interv	val: 1.5 cl	М			





Markers only explained ~5 % of variation

P. maxima QTL / GWAS analysis



Genomic selection (GS)



□ Most traits are complex involving many genes of small effect

- Need to simultaneously search for all genes of small effects
- □ GS application¹
 - Divide genome into many segments each with many loci
 - Estimate the genetic effect of each segment from a reference population (model training)
 - Genotype target population and sum segment effects to get genomic breeding values for individuals
- Benefits over traditional selective breeding
 - Increasing accuracy of selection (~0.7-0.8)
 - Reducing the generation interval
 - Allows within family selection
 - Minimizes inbreeding rates





Meuwissen et al, (2001)

P. maxima GS genetic resources



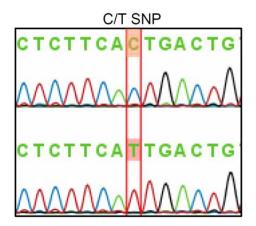
Genotype-by-Sequencing (GBS) SNPs for GS application

- ddRAD-tag, ~48,000 high quality SNPs identified
- SNPs and Indels separately identified
- High data integrity, >98% replicate repeatability
- \circ Genotype founders with 48K SNPs impute remainder
- Genotype remainder (10,000-15,000 animals) with
 5K SNPs @ \$20 each

Phenotypic / tissue data for GS application

- Grow-out commercial data 2003-2012 (> 500,000 animals, qualitative only)
- NBC commercial data 2013-ongoing (> 100,000 animals, qualitative + quantitative)

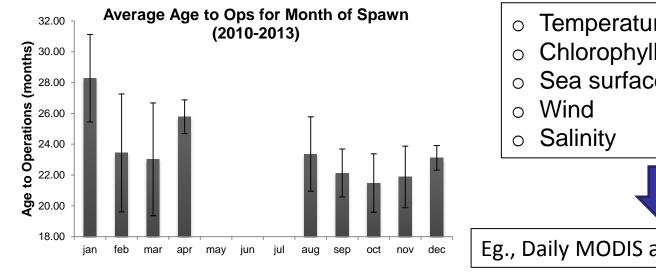
GS program commenced February 2015

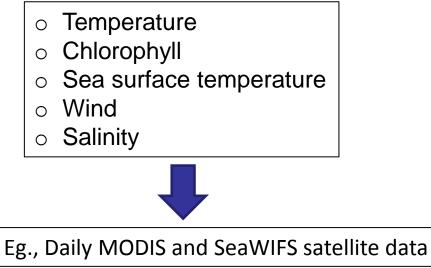


Issues Implementing Genomic selection



- Commercial operations separate host / donor lines
- Recording reliable phenotypic data (qual + quant)
- Environmental heterogeneity and seasonal variation





Month of the Year

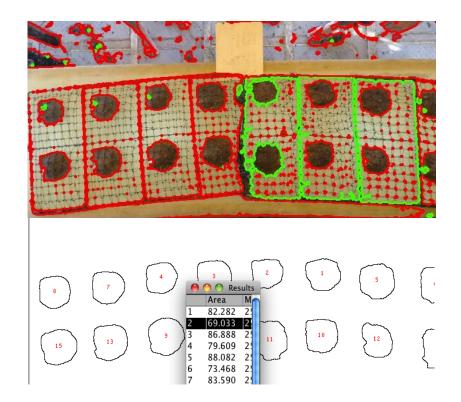
All are being measured to increase the accuracy of GS

Quantitative Phenotypic Data



- Host Oyster Growth automated with GoPro & image analysis
 - Routinely throughout entire production cycle development

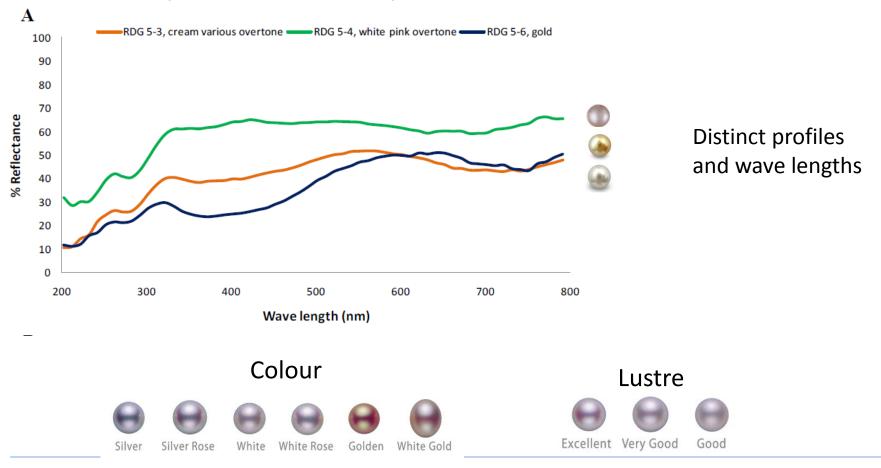




Quantitative Phenotypic Data



- Host Oyster Growth automated with GoPro & image analysis
- Donor Oyster Pearl Quality spectrophotometry



Final Comments



- Long-term 12+ year research program with industry
- Generated fundamental base-line data
- 1,000's genetic markers available at relative low cost (~\$20 animal)
- Adequate animal records / tissue available 10+ years
- Highly reliable phenotypic & environmental data most important
- Genomic Selection a real option for *P. maxima*, but need to integrate environmental variation effect on models